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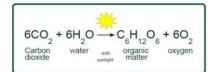


Size (Groups)

- Small cells warm and nutrient-poor conditions (stratified condition)
- Larger cells cold and nutrient-rich conditions

Nutritional strategy of phytoplankton

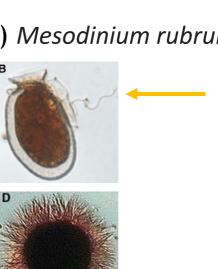
Autotrophs



Mixotrophs (combining photosynthesis and heterotrophy) Mesodinium rubrum

Vertical Movements

- Flagellate phytoplankton
- Mesodinium rubrum (hair-like structure)



Microphytoplankton

Nanophytoplankton

Picophytoplankton

Total Primary Producers

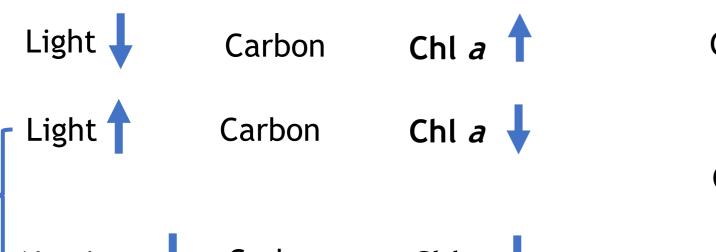
- 1. Chlorophyll a Concentration (Chl a, mg m⁻³)
- 2. Total Phytoplankton Biomass (C, µg C l-1)





Particulate backscattering coefficient (bbp), a proxy for particulate organic carbon (POC, carbon incorporated in living organisms e.g., phytoplankton, heterotrophic bacteria and nanoflagellates, detritus) from which phytoplankton biomass.

3. Carbon to Chlorophyll *a* ratio (C: Chl *a*)

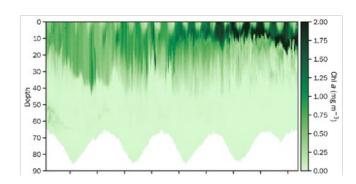


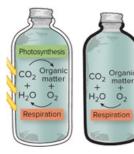
C: Chl a

Winter



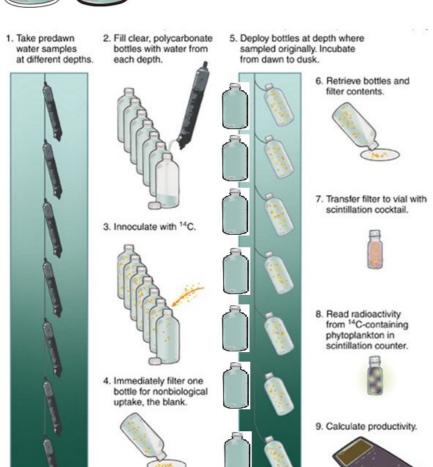
Summer



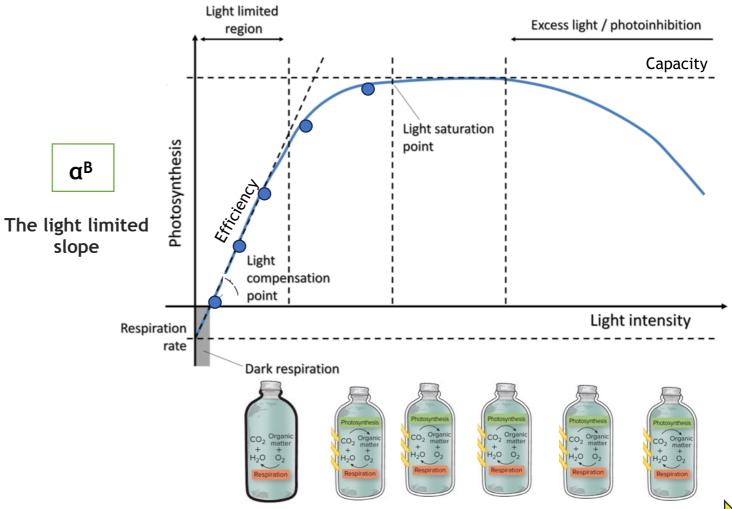


4. Primary Production

 ${\rm P^B}_{\rm max}$



The maximum photosynthetic rate



Primary Production Model (Platt et al., 1980)

$$PP_{\text{int}} = P^{B}_{max} \times \left(1 - e^{\alpha^{B} I_{0}^{m} / P^{B}_{max}}\right) \times \text{Chl } a$$

- **PP**_{int}: Integrated primary production (mg C m⁻² d⁻¹)
- P^B_{max}: The light-saturated maximum rates of the *P-E* curve normalized to chlorophyll *a* concentration
- α^B: The light-limited slope of the *P-E* curve normalized to chlorophyll *a* concentration
- I_o^m: Photosynthethic Activer Radiation (PAR) at depth
- Chl a: Chlorophyll a concentration at depth

- Vertically Generalized Production Model (VGPM, Behrenfeld and Falkowski, 1997),
- Carbon-based Productivity Model (CbPM, Behrenfeld et al., 2005)

Phytoplankton Succession

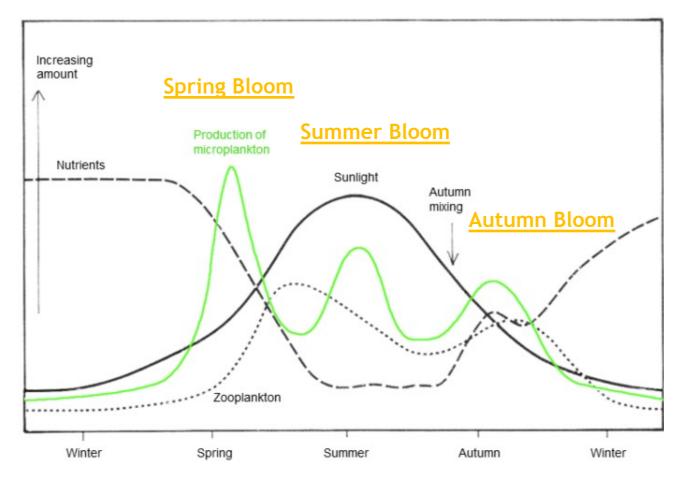


Fig. 1. Picture adapted from Rosenberg 1982 and Hällfors & Niemi 1986.

Summer Bloom

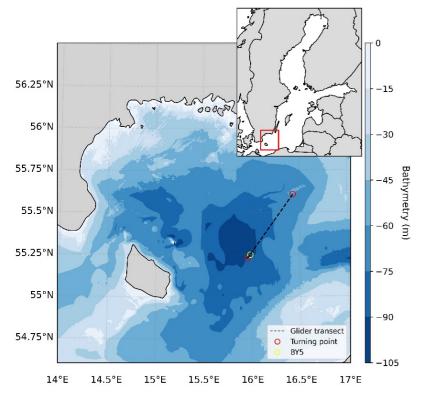


- 1. Produce toxins
- 2. Eutrophication
- 3. Contribute to oxygen depletion



3rd March 2021 and 15th February 2022

Study Area



Swedish Oceanographic Data Centre (SHARK) (0-10 m)



- Inorganic nutrients (Nitrate+Nitrite)
- 2. Chlorophyll a concentration (Chl a) \bigstar
- 3. Phytoplankton Biomass (pico-, nano- and microphytoplankton) 🤺
- 4. Photosynthetic parameters: light-saturated maximum rates (P^{B}_{max}) and light-limited slope (α^{B})
- 5. Primary Production 🕎





- 1. Salinity, temperature, and depth ——— Mixed layer depth (MLD)
- 2. Photosynthetically Active Radiation (PAR) Euphotic zone
- 3. Particulate Backscatter (700 nm), as a proxy of POC
- 4. Chlorophyll a concentration \star
- 5. Primary Production Model 솼





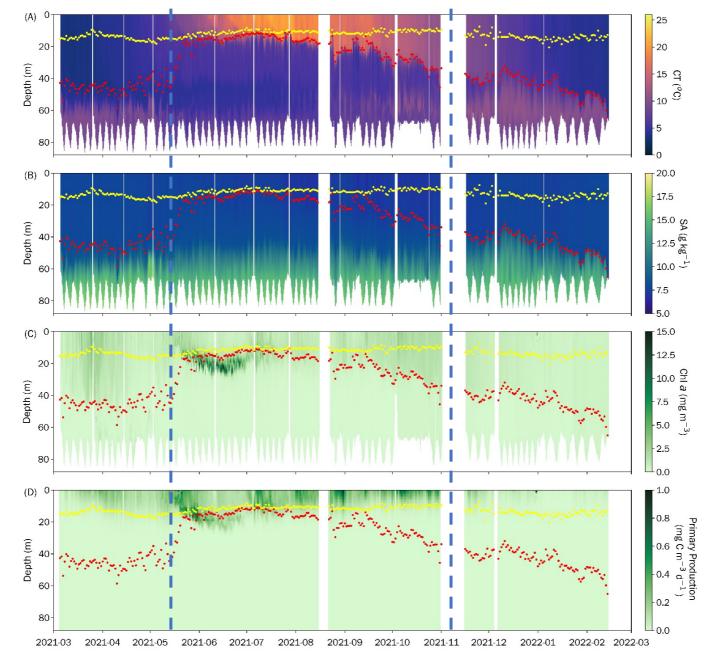
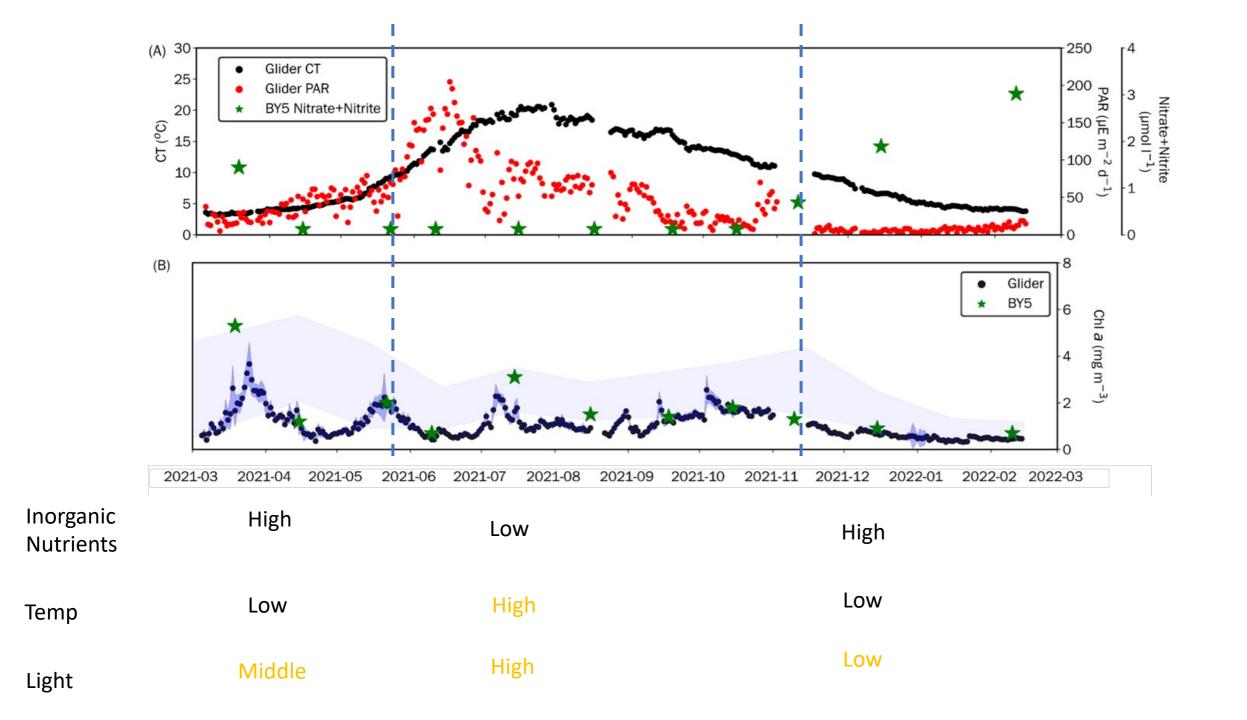


Fig. 2. Vertical variations of conservative temperature (CT) (A), absolute salinity (SA) (B), chlorophyll a concentration (Chl a) (C) and primary production (D) collected by the glider during the study period (2021/2022). The red and yellow dots represent the mixed layer depths (MLD) and euphotic depths, respectively.



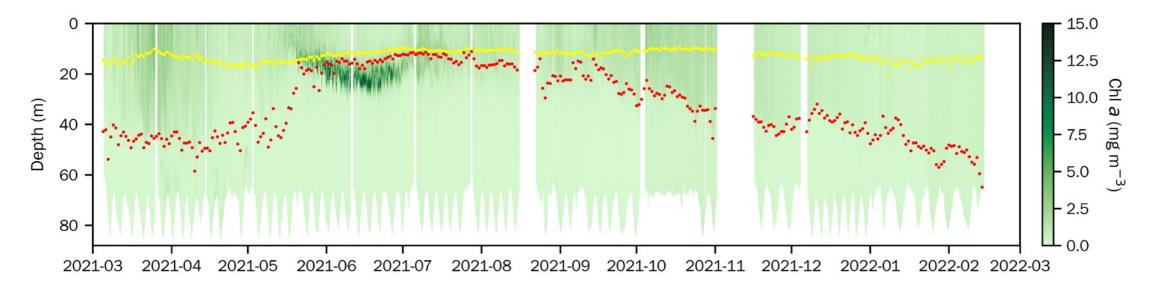
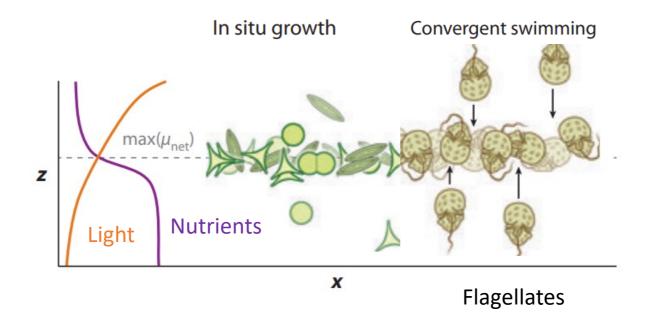


Fig.3. Vertical variations of chlorophyll *a* concentration (Chl *a*) collected by the glider during the study period (2021/2022). The red and yellow dots represent the mixed layer depths (MLD) and euphotic depths, respectively.



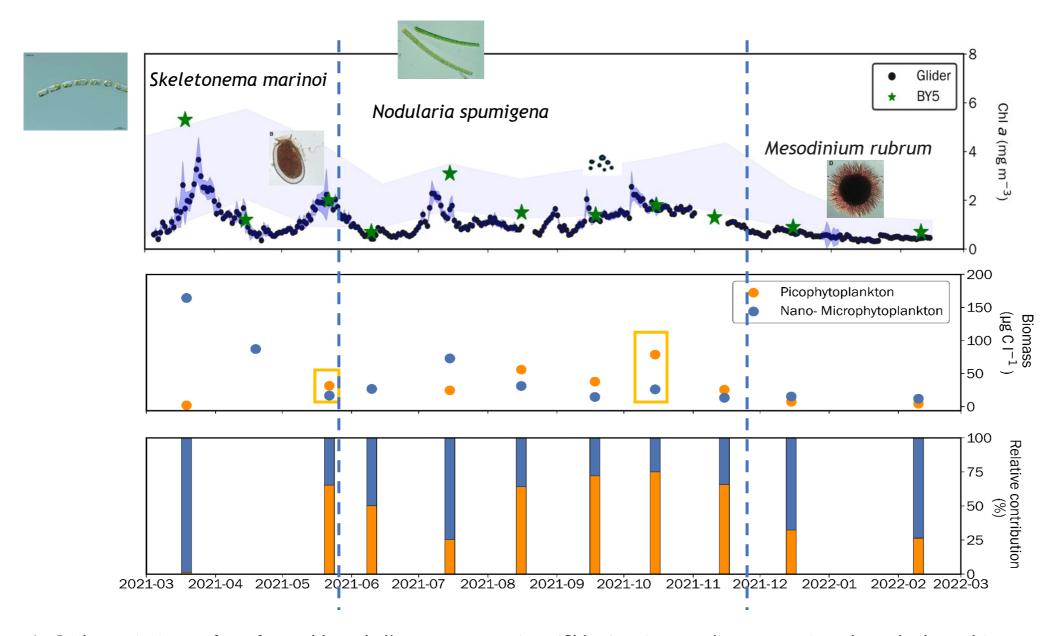


Fig. 4. Daily variations of surface chlorophyll a concentration (Chl a), pico- and nano-, microphytoplankton biomass and their relative contribution collected at BY5 station and by the glider during the study period (2021/2022). Light blue shaded area corresponds to the monthly average monitoring Chl $a \pm$ standard deviation.

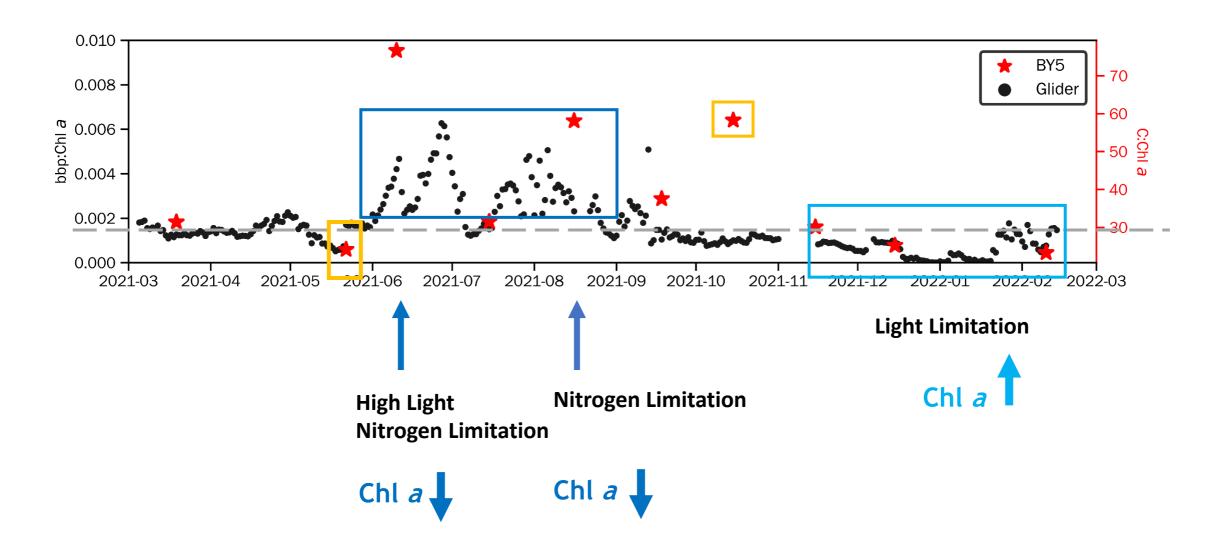


Fig. 5. Daily variations of C:Chl a ratio and bbp:Chl a ratio collected at BY5 station and by the glider during the study period (2021/2022).

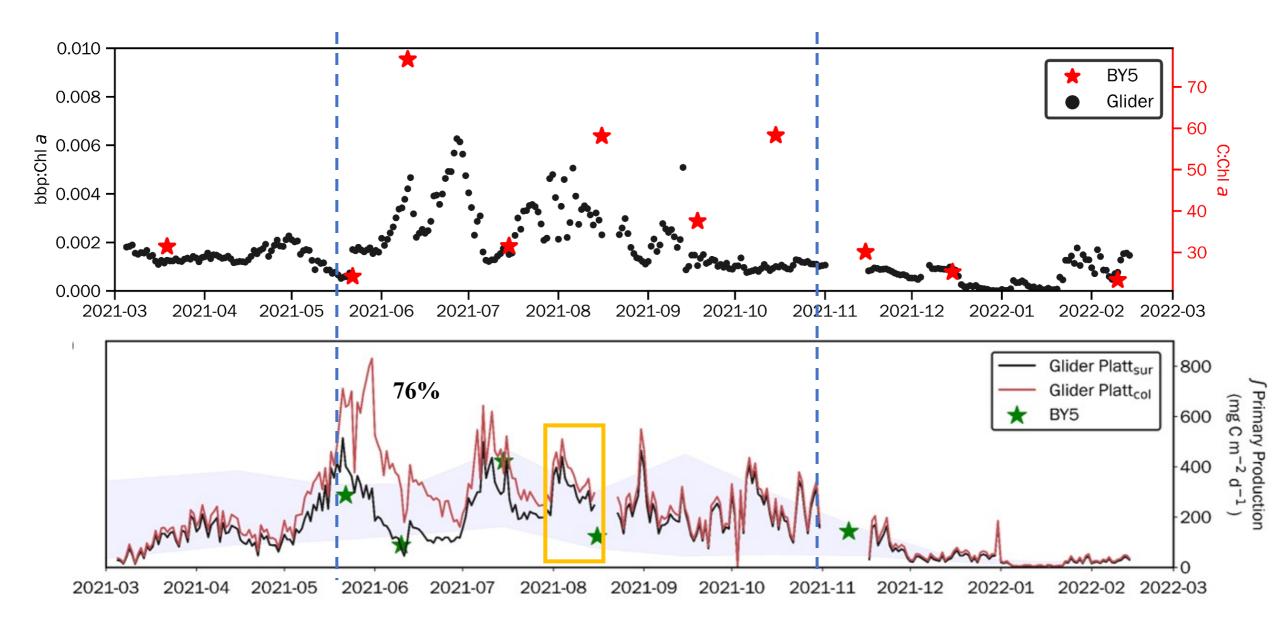


Fig. 6. Daily variations of bbp:Chl a and C:Chla ratio (A) and primary production (B) collected at BY5 station and estimated using the glider data during the study period (2021/2022).

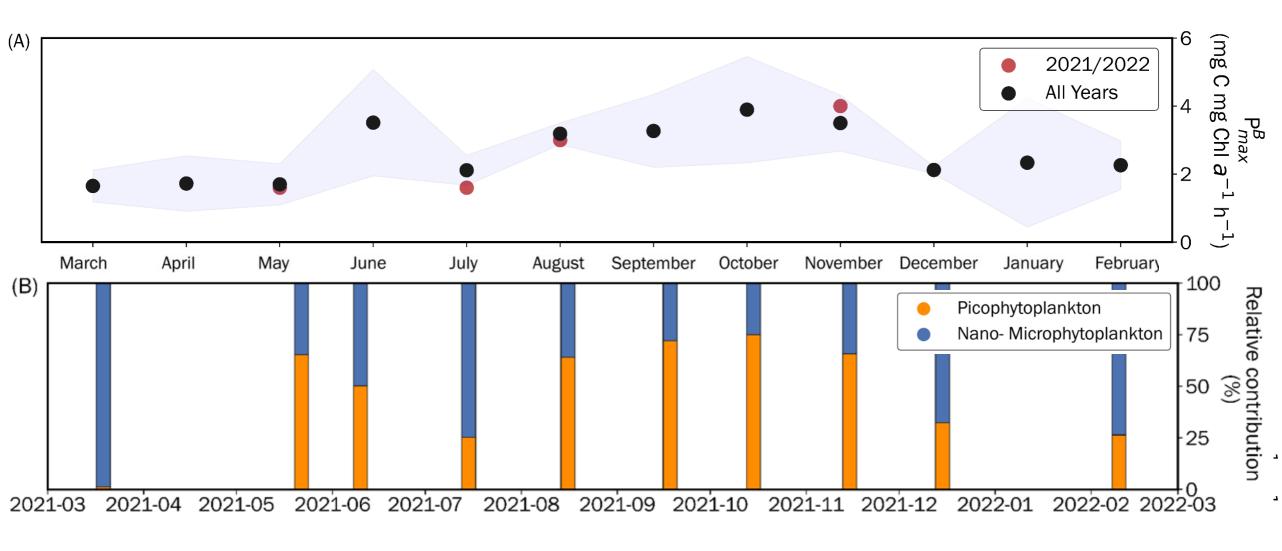


Fig. 7. Monthly variations of photosynthetic parameters: P^{B}_{max} collected at BY5 station during the study period (2021/2022) and the monitoring program (All Years) (A) and the relative contribution of pico- and nano-, microphytoplankton to the total phytoplankton biomass (B). Light blue shaded area corresponds to the monthly average monitoring photosynthetic parameter \pm standard deviation.

Conclusion:

- Gliders can fill the gap in Chlorophyll a and Primary Production measurements
- Deep Chlorophyll Maximum ——— Integrated Primary Production (up to 76% during the summer)
- Uncouple between Chlorophyll a concentration, Phytoplankton Biomass and Primary Production
- Picophytoplankton play a significant role in phytoplankton dynamics and primary production

Future work:

- Total Phytoplankton Biomass
- Explore other proxy from glider data to explain phytoplankton productivity (oxygen diel method)
- Expand the scope of this work to the other VOTO observatories